

NaveniFlex Cell GM

General guidelines

- Reaction volume depends on sample size and should be sufficient to cover the entire sample area. A volume of approximately 40 $\mu\text{l}/\text{cm}^2$ is recommended.
- Use best practices when pipetting to minimize reagent consumption.
- Thoroughly defrost all buffers at room temperature. Vortex well before use.
- Centrifuge vials before pipetting.
- Keep enzymes on ice or on a frozen cooling block.
- Vortex and spin down Enzymes 1 and 2 before use.
- Add enzymes right before adding the reaction mix to the sample.
- The recommended Navenibody working dilution is 1:40. Depending on target abundance, Navenibodies can be titrated down to 1:80.
- The NaveniFlex Cell GM kit can be combined with traditional immunofluorescence (IF) provided that all primary antibodies are either raised in different species or labelled with fluorophores different from the ones used in the kit.
- Buffer 2 is light-sensitive. Always keep it protected from light.
- Remove excess washing buffer from samples before adding the reagent mix.
- Do not allow slides/samples to dry.
- Preheat the humidity chamber before each step.
- Incubation times or assay temperatures other than those specified in the instructions for use may negatively impact results. Warm washing buffer is only required for step 4.4. Additional warm washing steps may lead to signal deterioration.
- As with any product derived from biological sources, proper handling procedures should be used.
- Wear appropriate personal protective equipment to avoid contact of reagents with eyes and skin.
- Unused solutions should be disposed of according to local regulations.
- The NaveniFlex Cell GM assay is also applicable for 96-well plates. For more information and protocol, please visit www.navinci.se/products.

Important:



Appropriate precautions should be taken to avoid antibody cross-contamination. Cross-contamination is the primary source of unspecific background due to the high sensitivity of the assay.

Avoid bulk washing methods when different primary antibody pairs are used, or when washing different technical controls.

Kit components

Box 1:

Storage: 4 to 8°C



| Material | Art.no | Amount |
|---------------------|-------------|-------------------|
| Block (1X) | NT.1.100.01 | 4 ml |
| Diluent 1 (1X) | NB.1.100.02 | 8 ml |
| Diluent 2 (1X) | NB.1.100.02 | 4 ml |
| Navenibody G1 (40X) | NB.1.100.18 | 100 μl |
| Navenibody M2 (40X) | NB.1.100.19 | 100 μl |

Box 2:

Storage: -25 to -15°C. Protect from light!



| Material | Art.no | Amount |
|------------------------------|----------------|-------------------|
| Buffer 1 (5X) | NB.2.100.17 | 800 μl |
| Enzyme 1 (40X) | NF.2.100.11 | 100 μl |
| Buffer 2 (5X) Red or Atto647 | NF.2.100.12/14 | 800 μl |
| Enzyme 2 (40X) | NF.2.100.15 | 100 μl |



For more information, or to place an order, visit www.navinci.se/products
 Email: contact@navinci.se

Instructions for use

1. Pre-treatment (reagents not provided)

- 1.1. Fix and permeabilize cells according to the immunofluorescence protocol optimized for the primary antibodies utilized in this assay.
- 1.2. Where applicable, use a hydrophobic barrier pen around the sample to prevent spillage and sample drying.
- 1.3. Prepare two bottles of 1X TBS-T*; pre-warm one bottle to 37°C together with the required number of staining jars (a warm wash is required after Navenibody incubation at Step 4.4)

*TBS-T (Tris-buffered saline supplemented with 0.05% Tween 20).

2. Blocking

- 2.1 Add **Block** to the entire sample area.
- 2.2 Incubate for 60 min at 37 °C in a preheated humidity chamber.

3. Primary antibody incubation

- 3.1. Use the provided **Diluent 1** to dilute your primary antibodies to working concentration. Start with a concentration optimized for/ recommended by the antibody vendor for ICC.
- 3.2. Decant **Block** and add a sufficient volume of the antibody working solution from step 3.1 to cover the sample area.
- 3.3. Incubate in a humidity chamber for 60 min at 37 °C, or at 4 °C overnight.
- 3.4. Decant the antibody solution, wash slides 2x 10 sec and 1x 15 min with 1X TBS-T in a staining jar under gentle agitation. Wash controls separately.

4. Navenibody incubation

- 4.1. Dilute **Navenibody G1** and **Navenibody M2** 1:40 in **Diluent 2**.
- 4.2. Add a sufficient volume of the Navenibody working solution from step 4.1 to cover the sample area.
- 4.3. Incubate in a preheated humidity chamber for 60 min at 37 °C.
- 4.4. Decant the Navenibody solution and wash slides with the **pre-warmed** 1X TBS-T from Step 1.3. Wash for 2x 10 sec and 1x 15 min in the pre-warmed staining jar under gentle agitation.

5. Reaction 1

- 5.1. Start preparing Reaction 1 by diluting **Buffer 1** 1:5 in water. Vortex and spin down.
- 5.2. Add **Enzyme 1** so that it is diluted 1:40. Mix gently by pipetting and spin down, see example calculation in Table 1 below.

Table 1. Example Calculation for a 40 µl reaction volume, sufficient for a 1 cm² area.

| Reagent for 1 cm ² sample area | Volume |
|---|--------|
| H ₂ O | 31 µl |
| Buffer 1 (5x) | 8 µl |
| Enzyme 1 (40x) | 1 µl |
| Total volume: | 40 µl |

- 5.3. Add a sufficient volume of Reaction 1 to cover the sample area.
- 5.4. Incubate in a preheated humidity chamber for 30 min at 37 °C.
- 5.5. Decant the solution, rinse once with 1 X TBS-T, then wash in 1x 5 min with 1X TBS-T in a staining jar under gentle agitation.

6. Reaction 2: protect from light!

- 6.1. Start preparing Reaction 2 by diluting **Buffer 2** 1:5 in water. Vortex and spin down.
- 6.2. Add **Enzyme 2** so that it is diluted 1:40. Mix gently by pipetting and spin down.
- 6.3. Add a sufficient volume of Reaction 2 to cover the sample area.
- 6.4. Incubate in a preheated humidity chamber for 90 min at 37 °C.
- 6.5¹. Decant the solution and wash slides for 2 min with 1X TBS in a staining jar under gentle agitation.

7. Nuclear stain and mounting (reagents not provided; protect from light!)

- 7.1. Remove excess washing buffer from the slides.
- 7.2. Add DAPI or a nuclear stain of your choice with a similar emission spectrum; dilute and incubate as recommended by the vendor.
- 7.3. Decant the solution, wash slides 2x 10 min in 1X TBS in a staining jar under gentle agitation.
- 7.4. Perform a final 15 min wash in 0.1X TBS in a staining jar under gentle agitation. Dry slides in a slide centrifuge or air-dry them, and mount slides with a coverslip using an antifade mounting medium.

8. Imaging

- 8.1. Image your slides with a fluorescence or confocal microscope using a 20x objective or higher.
- 8.2. For imaging, a filter set corresponding to DAPI, Cy3.5 and Cy5 is required. See table 2 below.

Table 2. Filter sets for imaging.

| Filter set | Detecting | λ Excitation | λ Emission |
|--------------------|--|--------------|------------|
| DAPI | Nuclei | | |
| Cy3.5/ TexasRed | Proximity signal detected by Buffer 2 Red | 585-595 nm | 615-625 nm |
| Cy5 | Proximity signal detected by Buffer 2 Atto647 | 635-645 nm | 665-675 nm |

¹(Optional) Fluorescently labelled primary antibodies may be diluted in diluent 1 and added after at this step if co-staining is desired. Dilute and incubate as recommended by the manufacturer.